This article was downloaded by: On: *29 January 2011* Access details: *Access Details: Free Access* Publisher *Taylor & Francis* Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Supramolecular Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713649759

Effect of the self-assembly of collagen on crystallisation of calcium carbonate in aqueous solution

Lin Yang^a; Feng Ye^a; Ruimin Xing^a; Baofang Zhang^a; Qiushi Ren^b

^a College of Chemistry and Environmental Science, Henan Normal University, Xinxiang, P.R. China ^b Institute for Laser Medicine and Bio-Photonics, Shanghai Jiao Tong University, Shanghai, P.R. China

To cite this Article Yang, Lin , Ye, Feng , Xing, Ruimin , Zhang, Baofang and Ren, Qiushi(2008) 'Effect of the self-assembly of collagen on crystallisation of calcium carbonate in aqueous solution', Supramolecular Chemistry, 20: 8, 761 – 763 **To link to this Article: DOI:** 10.1080/10610270802108936 **URL:** http://dx.doi.org/10.1080/10610270802108936

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doese should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



Effect of the self-assembly of collagen on crystallisation of calcium carbonate in aqueous solution

Lin Yang^a*, Feng Ye^a, Ruimin Xing^a, Baofang Zhang^a and Qiushi Ren^b

^aCollege of Chemistry and Environmental Science, Henan Normal University, Xinxiang, P.R. China; ^bInstitute for Laser Medicine and Bio-Photonics, Shanghai Jiao Tong University, Shanghai, P.R. China

(Received 6 September 2007; final version received 5 April 2008)

In order to investigate how the self-assembly of organic matrix influences crystallisation and growth of inorganic minerals, we selected collagen as the matrix and conducted three experiments of crystallisation of $CaCO_3$ in different reaction systems: H_2O system, as-assembled collagen fibrils system and self-assembling of collagen system. It is found that (i) the self-assembly process of organic matrix had a remarkable effect on the morphology of inorganic minerals: $CaCO_3$ crystals formed in the as-assembled collagen fibrils system were global clusters and those formed in the self-assembling of crystals: $CaCO_3$ crystals collagen system appeared as interlaced networks and (ii) the organic matrix decided the polymorph of crystals: $CaCO_3$ crystals were calcite in the H_2O system and appeared vaterite in the collagen system. From this study, we can conclude that the self-assembly of collagen fibrils greatly affect the crystallisation and growth of $CaCO_3$. Such results are significant in understanding the mechanism of biomineralisation in calcified tissues in general, and useful in the synthesis of biominerals.

Keywords: collagen fibrils; self-assembly; crystallisation; CaCO₃

Introduction

To this day, more than 60 different inorganic minerals in biological organisms are known. Some of them form mineralised tissues with extremely sophisticated shapes and highly organised microstructures (1, 2). From the studies of those minerals, it is found that crystal morphology, size, polymorph and orientation are affected by certain environmental conditions, in particular, the presence of a matrix (3-5). Therefore, some researchers have been intensively interested in the interactions between minerals and the matrix. Mann (6) proposed that the crystallisation of biominerals was a process similar to casting an inorganic replica in a static organic mould. Chang et al. (7) found that the self-assembled amelogenin protein microribbon structures controlled the oriented growth of apatite crystals during enamel mineralisation. However, the real biological organisms are sophisticated. The formation of inorganic minerals and the assembly of matrix are not two absolute independent processes (8). Generally, biomineralisation is concomitant with the selfassembly of matrix, and the latter controls the crystallisation tendency of the former. We studied the effect of self-assembling of sodium acrylate on the crystallisation of $CaCO_3$ (9), indicating that the self-assembly of sodium acrylate affected the shape of CaCO₃.

Experiment

In the present work, we studied the effect of selfassembling of collagen on the crystallisation of CaCO₃, which is more significant in understanding the mechanism of biomineralisation. CaCO₃, one of the most important building materials in biomineralising systems such as alga, trilobites, fish, mollusca, sea squirt, etc. makes up an attractive model mineral for studies in the laboratory (10, 11). Collagen is the most important and abundant structural protein for the extracellular matrix (12). Generally, there are several different types of collagen. We selected type I collagen as the matrix. Type I collagen monomer is stable below 4°C and pH \leq 2. When the temperature is 30°C and the pH value also reaches 5, the network fibrils can be formed by the self-assembly of the collagen monomer (13).

A set of equipments were specially designed. During the reaction process, the pH of the reaction solution (collagen + CaCl₂) was gradually increased by slow diffusion of NH₃ from the NH₄HCO₃ solution, as well as providing CO_3^{2-} for the reaction solution. The increase in the pH value induced the self-assembly of collagen monomer to network fibrils. The process of the self-assembly of collagen and crystallisation of calcium carbonate took place simultaneously.

ISSN 1061-0278 print/ISSN 1029-0478 online © 2008 Taylor & Francis DOI: 10.1080/10610270802108936 http://www.informaworld.com

^{*}Corresponding author. Email: yanglin1819@163.com

In a typical synthesis, three sets of experiments were carried out at 30°C in isolated environmental chambers. The first set was the experiment with the self-assembly of collagen monomer. The chamber contained two beakers one with 0.25 M NH₄HCO₃ solution and the other with a solution of pure collagen monomer (1.0 mg/ml). The second set was the control experiment, i.e. crystallisation of calcium carbonate in H₂O and as-assembled fibrils systems, respectively. The chamber contained three beakers - one with 0.25 M NH₄HCO₃ solution, another with CaCl₂ solution ($[CaCl_2] = 1.0 \text{ M}$, pH = 2) and the third one with CaCl₂ solution and as-assembled fibrils (1.0 M CaCl₂, 1.0 mg/ml collagen monomer). The third set was the master experiment with crystallisation of calcium carbonate during the process of self-assembling collagen monomer. The chamber contained two beakers - one with the 0.25 M NH₄HCO₃ solution and the other with a solution of CaCl₂ and collagen monomer (1.0 M CaCl₂, 1.0 mg/ml collagen monomer, pH = 2).

In order to comprehensively understand the selfassembly process of collagen monomer and the influence of the self-assembly collagen monomer to crystallisation of calcium carbonate, a set of time-series experiments in the third set were run (12, 24 and 72 h after sealing the system), respectively. The reaction solutions were withdrawn for transmission electron microscopy (TEM) observation, and the obtained solid products were characterised by powder X-ray diffraction (XRD) on a Rigaku Dmax-2000 diffractometer with Cu Kα radiation.

Discussion

To investigate the conditions under which collagen monomers self-assemble into fibrils, the TEM images of samples obtained at different reaction time of 12 h (pH 4.22) and 72h (pH 6.78), are presented in Figure 1(A) and (B), respectively. Figure 1(A) shows that there is a 1D strand-like

structure, in agreement with the original observation of TEM images of collagen monomer, indicating that the self-assembly process still did not happen when the pH value was 4.22. Figure 1(B) shows an infinite 3D network structure of collagen fibrils, indicating that network fibrils had been formed by the self-assembly of collagen monomer by increasing the pH value to 6.78 through diffusion of NH₃, when the self-assembly of collagen monomer proceeded for 72 h.

To contrast the differences of crystallisation of CaCO₃ in the pure H₂O and as-assembled fibrils systems, the TEM images and XRD patterns of samples were observed and are shown in Figure 2. The morphology of CaCO₃ (Figure 2(A)) formed in the pure H₂O system appeared a typical rhombohedral, and its XRD pattern (Figure 2(B)) proved that the crystal was calcite. The morphology of CaCO₃ formed in the as-assembled fibrils solution system (Figure 2(C)) were global clusters with a broader range in size, and its XRD pattern (Figure 2(D)) proved that the crystals were pure vaterite. It was obvious that the polymorph and morphology of calcium carbonate were relevant to the presence of a matrix.

Comparing Figure 2(C) with Figure 1(B), it is found that morphologies of vaterite $CaCO_3$ are spherical (Figure 2(C)), which are similar to the vacancies in the collagen fibrils structure (Figure 1(B)). It could be inferred that organic matrix as a scaffold-like template greatly influences the morphologies of CaCO₃.

In order to investigate the interaction between the selfassembly process of collagen monomer and crystallisation of CaCO₃, TEM analyses were performed after proceeding for different periods. Figure 3(A)-(C) displays the typical morphology of the samples for 12 h (pH 4.42), 24 h (pH 6.87) and 72h (pH 6.78), respectively. After proceeding for 12h (pH 4.42), the TEM image (Figure 3(A)) shows that the aggregation of crystals along the 1D direction was in good agreement with the structure of collagen monomer (Figure 1(A)), which could be attributed to the strand-like structure of monomer regulating the crystal growth. After 24 h, the one-dimensional arrays of the vaterite crystals had began to





Figure 1. TEM images of the self-assembly of pure collagen monomer to fibrils at different time: (A) 12h and (B) 72h, respectively.



Figure 2. (A) TEM image and (B) XRD pattern of the crystals formed in the pure H₂O system; (C) TEM image and (D) XRD pattern of the crystals formed in as-assembled fibrils system.



Figure 3. TEM images of crystals of calcium carbonate in the self-assembling of collagen monomer at different time: (A) 12 h, (B) 24 h and (C) 72 h, respectively.

intercross (Figure 3(B)), responding to the cross-linked fibrils. If the reaction proceeds to 72 h (pH 6.78), the morphology of crystals completely respond to the network structure of fibrils. Meanwhile, the result of its XRD pattern indicated that the crystals were vaterite.

In summary, in this work, the crystallisations of $CaCO_3$ were investigated in the pure H_2O system, the as-assembled collagen fibrils system and the process of self-assembling of collagen monomer, respectively. It is found that the process of assembly of organic matrix, collagen monomer, controlled the morphology of inorganic minerals $CaCO_3$, and the organic matrix influences the polymorph produced. Such experimental results are significant to understand the mechanism of biomineralisation and provide an illuminating synthetic approach to inorganic minerals with the complex form.

Acknowledgements

This work was supported by the National Basic Research Program of China (Grant no. 2005CB724306) and the National Science Foundation of China (Grant no. 20771036).

References

- (1) Lowenstam, H.A. Science 1981, 211, 1126–1131.
- (2) Xu, A. W.; Ma, Y.R.; Cölfen, H. J. Mater. Chem. 2007, 17, 415–449.
- (3) Addadi, L.; Weiner, S. Angew. Chem. 1992, 104, 159–176.
 (4) Bres, F.; Hutchinson, J.L. J. Biomed. Mater. Res. 2002,
- 63, 433-440.(5) Cölfan H: Antoniatti M Anagw Cham 2005 44
- (5) Cölfen, H.; Antonietti, M. Angew. Chem. 2005, 44, 5576–5591.
- (6) Mann, S. Biomineralization Principles and Concepts in Bioinorganic Materials Chemistry; Oxford University Press: Oxford, UK, 2001; Vol. 6, p 23.
- (7) Chang, D.; Giuseppe, F.; Fermani, S.; Abbott, C.; Moradian-Oldak, J. *Science* **2005**, *307*, 1450.
- (8) Hartgerink, J.D.; Beniash, E.; Stupp, S.I. Science 2001, 23, 1684.
- (9) Guo, Y.M.; Yang, L.; Liao, Z.J.; Zhang, X.Y.; Zhu, S.F.; Jiang, K. *Macromol. Biosci.* **2003**, *3*, 163.
- (10) Berman, A.; Addadi, L.; Weiner, S. *Nature* **1988**, *311*, 546–548.
- (11) Addadi, L.; Raz, S.; Weiner, S. Adv. Mater. 2003, 15, 959–970.
- (12) Nimni, M.E., Ed. *Biochemistry*; CRC Press: Boca Raton, FL, 1988; Vol. I.
- (13) Goh, M.C.; Paige, M.F.; Gale, M.A.; Yadegari, I.; Edirisinghe, M. J. Strzelczyk 1997, 239, 95–102.